

FORM PTO-1390
(REV. 9-2001)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371

ATTORNEY'S DOCKET NUMBER

YAM 2 0014

U.S. APPLICATION NO (If known, see 37 CFR 1.5

10/030298

INTERNATIONAL APPLICATION NO.
PCT/JP00/04167INTERNATIONAL FILING DATE
23 June 2000PRIORITY DATE CLAIMED
23 June 1999TITLE OF INVENTION
DRUG FOR INHIBITING MYOMETRIAL CONTRACTIONAPPLICANT(S) FOR DO/EO/US
YANAGITA, Toshihiko

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (21) indicated below.
4. The US has been elected by the expiration of 19 months from the priority date (Article 31).
5. A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. is attached hereto (required only if not communicated by the International Bureau).
 - b. has been communicated by the International Bureau.
 - c. is not required, as the application was filed in the United States Receiving Office (RO/US).
6. An English language translation of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. is attached hereto.
 - b. has been previously submitted under 35 U.S.C. 154(d)(4).
7. Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. are attached hereto (required only if not communicated by the International Bureau).
 - b. have been communicated by the International Bureau.
 - c. have not been made; however, the time limit for making such amendments has NOT expired.
 - d. have not been made and will not be made.
8. An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. An English language translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11 to 20 below concern document(s) or information included:

11. An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. A FIRST preliminary amendment.
14. A SECOND or SUBSEQUENT preliminary amendment.
15. A substitute specification.
16. A change of power of attorney and/or address letter.
17. A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825.
18. A second copy of the published international application under 35 U.S.C. 154(d)(4).
19. A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).
20. Other items or information: Application Data Sheet
Sequence Listing (12 pages)
Verified Statement Under 37 CFR 1.821(f)

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December 21, 2001
I hereby certify that this paper is being transmitted via
U.S. Postal Service "Express Mail, Post 6 days to addressee" service
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Assistant Commissioner for Patents, Washington, D.C. 20231.

LAURIE A. BOYLAN

(TYPED OR PRINTED NAME OF SENDER)

Laurie A. Boylan

U.S. APPLICATION NO (if known, see 37 CFR 1.5)	INTERNATIONAL APPLICATION NO	PCT/JP00/04167	ATTORNEY'S DOCKET NUMBER YAM 2 0014
10/030298		CALCULATIONS PTO USE ONLY	
<p><input checked="" type="checkbox"/> The following fees are submitted:</p> <p>BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)):</p> <p>Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO \$1040.00</p> <p>International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$890.00</p> <p>International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$740.00</p> <p>International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$710.00</p> <p>International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) \$100.00</p>			
ENTER APPROPRIATE BASIC FEE AMOUNT =		\$890.00	
<p>Surcharge of \$130.00 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e)).</p>		\$	
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE
Total claims	17 - 20 =	0	x \$18.00
Independent claims	3 - 3 =	0	x \$84.00
MULTIPLE DEPENDENT CLAIM(S) (if applicable)		+ \$280.00	
TOTAL OF ABOVE CALCULATIONS =		\$ 890.00	
<input type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.		+	
SUBTOTAL =		\$ 890.00	
<p>Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)).</p>		\$	
TOTAL NATIONAL FEE =		\$ 890.00	
<p>Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property</p>		+ \$ 40.00	
TOTAL FEES ENCLOSED =		\$ 930.00	
		Amount to be refunded:	\$
		charged:	\$
<p>a. <input checked="" type="checkbox"/> A check in the amount of \$ <u>930.00</u> to cover the above fees is enclosed.</p> <p>b. <input type="checkbox"/> Please charge my Deposit Account No. _____ in the amount of \$ _____ to cover the above fees. A duplicate copy of this sheet is enclosed.</p> <p>c. <input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. <u>06-0308A</u> duplicate copy of this sheet is enclosed.</p> <p>d. <input type="checkbox"/> Fees are to be charged to a credit card. WARNING: Information on this form may become public. Credit card information should not be included on this form. Provide credit card information and authorization on PTO-2038.</p>			
<p>NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137 (a) or (b)) must be filed and granted to restore the application to pending status.</p>			
<p>SEND ALL CORRESPONDENCE TO: Richard M. Klein FAY, SHARPE, FAGAN, MINNICH & McKEE, LLP 1100 Superior Avenue, Seventh Floor Cleveland, OH 44114 RKlein@faysharpe.com (216) 861-5582</p>			
 SIGNATURE <hr/> <p>Richard M. Klein</p> <hr/> <p>NAME</p> <hr/> <p>33,000</p> <hr/> <p>REGISTRATION NUMBER</p>			

10/030298

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JC13 Rec'd PCT/PTO 21 DEC 2001

I hereby certify that this Preliminary Amendment is being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 CFR 1.10 on the date indicated above and is addressed to: BOX Patent Application, Assistant Commissioner of Patents, Washington, D.C. 20231.

Laurie A. Boylan
By: Laurie A. Boylan

#31
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PATENT

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of)
YANAGITA, Toshihiko)
International Application No. PCT/JP00/04167)
International Filing Date June 23, 2000)
For: DRUG FOR INHIBITING MYOMETRIAL)
CONTRACTION)
Attorney Docket No. YAM 2 0014)

Cleveland, Ohio 44114
December 21, 2001

PRELIMINARY AMENDMENT

BOX Patent Application
Assistant Commissioner for Patents
Washington, D.C. 20231

Dear Sir:

Prior to the examination of the merits and/or calculation of the fees, kindly amend the above-identified application as follows:

IN THE CLAIMS:

Please amend claims 3-7 and 11-13 to read as follows:

3. (Amended) A composition according to claim 1, used to prevent premature labor.

4. (Amended) A composition according to claim 1, used to prevent miscarriage.

5. (Amended) A composition according to claim 1, used to arrest parturition prior to cesarean section.
6. (Amended) A composition according to claim 1, used to treat dysmenorrhea.
7. (Amended) A composition according to claim 1, wherein the adrenomedullin is:
- (a) a peptide comprising an amino acid sequence from Ser in position 13 to Tyr in position 52 of SEQ ID NO: 2 in SEQUENCE LISTING; or
- (b) a peptide comprising an amino acid sequence having one or several amino acid deleted, substituted, or added in the amino acid sequence (a), and having an action of inhibiting myometrial contraction.
11. (Amended) A composition according to claim 1, wherein the C-terminus of the adrenomedullin is amidated.
12. (Amended) A composition according to claim 1, wherein Gly is added to the C-terminus of the adrenomedullin.
13. (Amended) A composition according to claim 1, wherein in the adrenomedullin, Cys in position 16 and Cys in position 21 of SEQ ID NO: 2 in SEQUENCE LISTING are crosslinked.

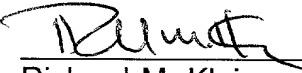
REMARKS

Entry of this preliminary amendment is requested at the U.S. Patent and Trademark Office's earliest convenience.

Respectfully submitted,

FAY, SHARPE, FAGAN,
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By:


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Attachment - Version With Markings to Show Changes Made

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VERSION WITH MARKINGS TO SHOW CHANGES MADE

Please amend claims **3-7 and 11-13** (i.e. additions are underlined and deletions are bracketed) to read as follows:

IN THE CLAIMS:

3. (Amended) A composition according to claim 1 [or 2], used to prevent premature labor.
4. (Amended) A composition according to claim 1 [or 2], used to prevent miscarriage.
5. (Amended) A composition according to claim 1 [or 2], used to arrest parturition prior to cesarean section.
6. (Amended) A composition according to claim 1 [or 2], used to treat dysmenorrhea.
7. (Amended) A composition according to claim 1 [or 2], wherein the adrenomedullin is:
 - (a) a peptide comprising an amino acid sequence from Ser in position 13 to Tyr in position 52 of SEQ ID NO: 2 in SEQUENCE LISTING; or
 - (b) a peptide comprising an amino acid sequence having one or several amino acid deleted, substituted, or added in the amino acid sequence (a), and having an action of inhibiting myometrial contraction.
11. (Amended) A composition according to claim 1 [or 2], wherein the C-terminus of the adrenomedullin is amidated.
12. (Amended) A composition according to claim 1 [or 2], wherein Gly is added to the C-terminus of the adrenomedullin.

13. (Amended) A composition according to claim 1 [or 2], wherein in the adrenomedullin, Cys in position 16 and Cys in position 21 of SEQ ID NO: 2 in SEQUENCE LISTING are crosslinked.

10/030298

JC13 Rec'd PCT/PTO 21 DEC 2001

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SO043

DESCRIPTION

DRUG FOR INHIBITING MYOMETRIAL CONTRACTION

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TECHNICAL FIELD

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The present invention relates to a composition for inhibiting spontaneous myometrial contraction or bradykinin-induced contraction, comprising adrenomedullin. More specifically, the present invention relates to a composition for selectively inhibiting spontaneous myometrial contraction or bradykinin-induced contraction, comprising adrenomedullin.

15

BACKGROUND ART

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The management of premature labor is one of the most important issues in the field of obsterics. Premature labor refers to parturition on or after 22 weeks and before 37 weeks of pregnancy, which accounts for 5 to 10% of the total number of childbirths. A neonate delivered by premature labor is called a pronatus, which is frequently a low birth weight infant. Recently, although the care of neonates has progressed significantly, the morbidity and mortality of pronatases are higher compared to normally delivered neonates. Therefore, it is desired that premature labor is avoided to continue pregnancy as long as possible.

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At present, widely used drugs for preventing premature labor include β_2 -adrenergic sympathetic agonist, magnesium sulfate, and indomethacin (a prostaglandin synthesis inhibitor).

5 Ritodrine, which is a representative β_2 -adrenergic agonist, causes various cardiovascular and metabolic side effects, including tachycardia, elevation of renin secretion, and hyperglycemia in mothers (and hypoglycemia in neonates). Other β_2 -adrenergic agents, such as for example terbutaline and albuterol, cause side effects similar to those of ritodrine.

10 Magnesium sulfate having a plasma concentration of 4 to 8 mg/dL, which exceeds the therapeutically acceptable range, causes inhibition of cardiac conduction and neuromuscular transmission, hypopnea, and cardiac arrest. Therefore, when renal function is impaired, this agent is not preferable.

15 Indomethacin causes side effects in fetuses, such as for example pulmonary artery hypertension and persistent truncus arteriosus, which are contraindications to large dosage and long-term use of indomethacin.

20 As described above, currently known drugs for preventing premature labor have various drawbacks. Therefore, there is a demand for a novel drug for preventing premature labor without such drawbacks.

25 The mechanism of the onset of parturition, i.e., the beginning of pains, has not been yet fully revealed, but it has been suggested that oxytocin, prostaglandin, and the like having a uterine contracting action are involved. Bradykinin has a uterine contracting action similar to oxytocin and prostaglandin, but the physiological or pathophysiological meaning is still unknown. However, bradykinin is inherently an inflammatory mediator, and it

has been suggested that there is a possibility that premature labor and miscarriage is caused by abnormally increased bradykinin in the gravid uterus, (Reference 1; List of references are described at the end of this specification).
5 Therefore, if a drug capable of inhibiting spontaneous myometrial contraction or bradykinin-induced myometrial contraction action, or a drug capable of selectively inhibiting spontaneous myometrial contraction or bradykinin-induced contraction action, was especially found,
10 the drug would be expected to be useful not only for preventing premature labor but also for preventing miscarriage and arresting of parturition prior to cesarean section.

15 Further, such an agent would be expected to be useful in the treatment of dysmenorrhea. This is because dysmenorrhea is characterized by a periodic pain in association with menses during a ovulocycle, and the pain is believed to be derived from uterine contraction and
20 ischemia.

25 Adrenomedullin (AM), a member of the calcitonin gene-related peptide (CGRP) family, was originally isolated from human pheochromocytoma as a peptide having a hypotensive action (Reference 2). It is known that AM plays multiple roles in a variety of tissues (Reference 3). This suggests that AM has a nonuniform mechanism of action in an organism.

30 Levels of AM protein or AM mRNA in the female reproductive system (e.g., pituitary posterior lobe [Reference 3] and the uterus [Reference 4]), are as high as those in adrenal medulla. Also, level of circulating AM

in maternal blood (Reference 5), and AM and AM mRNA abundance in fetal-placental tissues (Reference 6) and the uterus (Reference 7) were both elevated during normal pregnancy. In gestosis, a complication of pregnancy, the maternal plasma AM level did not alter (Reference 5), or lowered (Reference 8), whereas the AM content in amniotic fluid and umbilical vein were higher, compared to normal pregnancy (Reference 9). However, physiological roles of AM and details of AM function in these fetal and maternal tissues remain elusive.

As to the effect of AM on uterine contraction, it has only been reported that AM inhibited galanin-induced tonic contraction of the uterus at high concentrations of 5 μ M or more, and the action of AM was eliminated by CGRP[8-37] (Reference 7). Galanin is a neuropeptide contained in a neuron (CGRP). However, the importance of galanin-induced uterine contraction is not understood at all. AM action is not found at concentrations of the order of nanomolar (nM) or less (which are the concentrations reported in a number of papers as concentrations at which AM can function). It has been only confirmed that AM can function at high concentrations of the order of micromolar (μ M) or more. Therefore, it is hardly believed that these effects reflect a physiological function of AM.

The motility of the uterus (contraction/relaxation) is not only regulated by nerves, such as sympathetic nerves and parasympathetic nerves, but also is coordinately regulated by various substances, such as for example CGRP (Reference 10), nitric oxide (NO), oxytocin, and prostaglandin F_{2 α} (PGF_{2 α} : a representative prostaglandin having various actions: elevation of blood

pressure; vasoconstriction; promotion of intestine movement; uterine contraction; promotion of regression of corpus luteum; and bronchoconstriction, and used as a parturifacient). Similar to bradykinin described above, substances which can cause abnormal contraction leading to premature labor also influence the motility of the uterus. However, what effect is obtained by AM on spontaneous uterine contraction; the contractility evoked by regulatory factors, such as oxytocin, PGF_{2α} and the like; or bradykinin-induced contraction, is not known at all.

The present invention is intended to solve the above-described problems. The objective of the present invention is to provide a novel agent for inhibiting, preferably selectively inhibiting, spontaneous myometrial contraction or bradykinin-induced contraction.

DISCLOSURE OF THE INVENTION

The inventor found that adrenomedullin, originally identified as a peptide having a hypotensive action, has an action of selectively inhibiting spontaneous myometrial contraction and bradykinin-induced contraction and that the inhibitor action is selective, and based on this finding, completed the present invention.

A composition of the present invention for inhibiting spontaneous myometrial contraction or bradykinin-induced contraction comprises AM. The composition of the present invention may be used for selectively inhibiting spontaneous myometrial contraction or bradykinin-induced contraction, preventing premature labor, preventing miscarriage, arresting parturition prior

to cesarean section, and treating dysmenorrhea.

In one embodiment, the AM includes: (a) a petptide comprising an amino acid sequence from Ser in position 13 to Tyr in position 52 of SEQ ID NO: 2 in SEQUENCE LISTING; (b) a peptide comprising an amino acid sequence having one or several amino acid deleted, substituted, or added in the amino acid sequence (a), and having an action of inhibiting myometrial contraction; (c) a petptide comprising an amino acid sequence from Tyr in position 1 to Tyr in position 52 of SEQ ID NO: 2 in SEQUENCE LISTING; (d) a peptide comprising an amino acid sequence having one or several amino acid deleted, substituted, or added in the amino acid sequence (c), and having an action of inhibiting myometrial contraction; (e) a petptide comprising an amino acid sequence from Ala in position -73 to Tyr in position 52 of SEQ ID NO: 2 in SEQUENCE LISTING; (f) a peptide comprising an amino acid sequence having one or several amino acid deleted, substituted, or added in the amino acid sequence (e), and having an action of inhibiting myometrial contraction; (g) a petptide comprising an amino acid sequence from Met in position -94 to Leu in position 91 of SEQ ID NO: 2 in SEQUENCE LISTING; and (h) a peptide comprising an amino acid sequence having one or several amino acid deleted, substituted, or added in the amino acid sequence (g), and having an action of inhibiting myometrial contraction.

In another embodiment, the C-terminus of the AM may be amidated. Gly may be added to the C-terminus of the AM.

In another embodiment, in the AM, Cys in position 16 and Cys in position 21 of SEQ ID NO: 2 in SEQUENCE LISTING

may be crosslinked. The crosslink may be a disulfide bond or a -CH₂-CH₂- bond.

5 A method of the present invention for preventing premature labor or miscarriage uses a composition comprising AM.

10 The present invention also provides use of AM in production of a drug for preventing premature labor or miscarriage.

BRIEF DESCRIPTION OF THE DRAWINGS

15 Figure 1(A) is a schematic diagram showing a uterus site from which uterine strips were isolated in examples. Figure 1(B) is a schematic diagram showing the shape of the prepared uterine strips.

20 Figure 2(a) is a graph showing the result of measuring myometrial contraction when distilled water was added to the uterine strip **a** in Figure 1(A). Figure 2(b) is a graph showing the result of measuring myometrial contraction when distilled water was added to the uterine strip **b** in Figure 1(A). Figure 2(c) is a graph showing the 25 result of measuring myometrial contraction when 100 nM AM was added to the uterine strip **c** in Figure 1(A). Figure 2(d) is a graph showing the result of measuring myometrial contraction when 100 nM AM was added to the uterine strip **d** in Figure 1(A).

30 Figure 3(a) is a graph showing the result of measuring myometrial contraction when 1 to 100 nM AM was added to the uterine strip **b** in Figure 1(A). Figure 3(b)

5 is a graph showing the result of measuring myometrial contraction when 1 to 100 nM AM was added to the uterine strip **c** in Figure 1(A). Figure 3(c) is a graph showing the result of measuring myometrial contraction when distilled water was added to the uterine strip **d** in Figure 1(A).

10 Figure 4(a) is a graph showing the result of measuring myometrial contraction when bradykinin and then 100 nM AM were added to the uterine strip **b** in Figure 1(A).

15 Figure 4(b) is a graph showing the result of measuring myometrial contraction when bradykinin and then distilled water were added to the uterine strip **b** in Figure 1(A).

15 Figure 5 is a diagram showing the amino acid sequence of AM derived from human pheochromocytoma. RE1 to RE6 indicate fragments produced by digesting the amino acid sequence with arginylendopeptidase.

20 Figure 6 is a diagram showing concentration-dependent inhibition of spontaneous myometrial contraction of the uterus by AM; and prevention by AM[22-52] or CGRP[8-37]. Figures 6(a) to 6(e) are representative records from five separate experiments having similar results. * $p<0.05$, comparison with responses without a drug (one-way analysis of variance); * $p<0.05$, comparision with AM alone (two-way analysis of variance).

30 Figure 7 is a diagram showing inhibition of bradykinin-induced uterine contraction which was caused by AM but not by prevention of oxytocin or PGF_{2α} by AM[22-52] or CGRP[8-37]. Figures 7(a) to 7(e) show representative records from five separate experiments having similar results.

BEST MODE FOR CARRYING OUT THE INVENTION

In embodiments of the present invention, protein purification and analysis methods, recombinant DNA techniques, and assays, which are known in the art, are employed unless otherwise specified.

I. Definition

Hereinafter, the terms used herein to explain the present invention will be described.

As described above, "adrenomedullin" is a peptide having a hypotensive action, originally isolated from human pheochromocytoma. The term "adrenomedullin" as used herein is not limited to the particular peptide, but includes peptides having substantial homology with the amino acid sequence of that peptide. Examples of the homologous peptides include species mutants and allelic mutants. Human-derived AM comprises an amino acid sequence from Tyr in position 1 to Tyr in position 52 of SEQ ID NO: 2 in SEQUENCE LISTING. (The peptide consisting of an amino acid sequence from Met in position -94 to Leu in position 91 of SEQ ID NO: 2 in SEQUENCE LISTING is believed to be preproadrenomedullin. The peptide obtained by processing of a signal peptide and consisting of an amino acid sequence from Ala in position -73 to Leu in position 91 of SEQ ID NO: 2 in SEQUENCE LISTING is believed to be proadrenomedullin. The peptide consisting of an amino acid sequence from Ser in position 13 to Tyr in position 52 of SEQ ID NO: 2 in SEQUENCE LISTING is an AM fragment which has been confirmed to have a hypotensive action. AM in any of the above-described forms may be employed in the present

invention.) Human-derived AM may be encoded by a polynucleotide sequence from T in position 447 to C in position 602 of SEQ ID NO: 1 in SEQUENCE LISTING. Porcine-derived AM comprises an amino acid sequence from Tyr in position 1 to Tyr in position 52 of SEQ ID NO: 4 in SEQUENCE LISTING. Porcine-derived AM may be encoded by a polynucleotide sequence from T in position 430 to C in position 585 of SEQ ID NO: 3 in SEQUENCE LISTING. Rat-derived AM comprises an amino acid sequence from Tyr in position 1 to Tyr in position 50 of SEQ ID NO: 6 in SEQUENCE LISTING. Rat-derived AM may be encoded by a polynucleotide sequence from T in position 433 to T in position 582 of SEQ ID NO: 5 in SEQUENCE LISTING.

Clearly, human-derived peptides are preferable for human diseases or treatment of a human. However, homologous peptides derived from other mammals may also be employed for some purposes. Further, comparison of human-derived peptides with peptide derived from other mammals is important when an attempt is made to obtain a variant maintaining a desired activity of a human-derived peptide.

AM used in the present invention is not necessarily limited to the above-described sequences, but includes, as subjects, homologous peptides having an amino acid sequence having one or several amino acid deleted, substituted, or added in the above-described sequences and maintaining a desired activity.

Amino acid conservative substitution is one preferable means for obtaining homologous peptides. Conservative substitution representatively includes substitutions conducted within the following groups:

glycine, alanine; valine, isoleucine, leucine; aspartic acid, glutamic acid; asparagine, glutamine; serine, threonine; lysine, arginine; and phenylalanine, tyrosine.

5 The homology between two amino acid sequences is determined by optionally introducing a gap to optimize residue matching. A peptide having an amino acid sequence, which has substantially homology with the amino acid sequence of human AM, has representatively about 60% homology with the amino acid sequence of human AM, preferably at least about 70%, more preferably at least about 80%, and in an especially preferable embodiment, at least about 90% or more. Software for determining homology is easily available.

15 In the present invention, a peptide is by definition referred to "having an action of inhibiting myometrial contraction" if the degree of spontaneous contraction of uterine muscle is about 90% or less and preferably about 80% or less of the value indicated in the Control sample of Example 1 or if the degree of bradykinin-induced contraction is about 90% or less and preferably about 80% or less of the value indicated in the Control sample of Example 2, when both are measured under substantially the same conditions as those of Example 1 below (the concentration of AM added was 100 nM).

20 In the present invention, a peptide is by definition referred to as "not inhibiting contraction" if the degree of spontaneous contraction of uterine muscle is about 90% or more and preferably about 95% or more of the value indicated in Control sample of in Example 1; if the degree of bradykinin-induced contraction is about 90% and

5 preferably about 95% or more of the value indicated in the Control sample shown in Example 2; or if degree of contraction induced by oxytocin or prostaglandin F_{2α} is higher about 90% and preferably about 95% or more of a value indicated before the addition of AM in Example 4, when measured respectively under substantially the same conditions as those of Example 1 (the concentration of AM added was 100 nM).

10 In the present invention, the peptide is referred to as "having an action of selectively inhibiting myometrial contraction" when although spontaneous myometrial contraction or bradykinin-induced contraction is inhibited, myometrial contraction induced by oxytocin and prostaglandin F_{2α} is not inhibited.

15

20 The C-terminus of a peptide used in the present invention may or may not be amidated. "Amidation of C-terminus" refers to one of modification reactions of a peptide, in which the COOH group of the C-terminal amino acid of a peptide is changed to the form of CONH₂. A number of biologically active peptides functioning in vivo are first biosynthesized as a precursor protein having a larger molecular weight. The precursor protein is then matured by 25 a modification reaction such as for example the amidation of the C-terminus. The amidation is conducted by a C-terminal amidating enzyme acting on the precursor protein. The precursor protein always includes a Gly residue on the C-terminal side of a residue to be amidated, which is frequently followed by a basic amino acid sequence pair, 30 such as for example Lys-Arg or Arg-Arg, on the C-terminal side (Reference 11).

II. AM having an action of inhibiting myometrial contraction

In the present invention, AM is used as an effective component of a composition for inhibiting, preferably selectively inhibiting, spontaneous myometrial condition or bradykinin-induced contraction. AM is also used as an effective component for producing a drug for preventing premature labor or miscarriage. AM may be those isolated from naturally-occurring sources, those produced using recombinant DNA techniques, or those chemically synthesized.

When AM is isolated from naturally-occurring sources, purification may be conducted, for example, in the following way. For example, firstly human pheochromocytoma is pulverized to obtain a crude extract, followed by various chromatography techniques so that adenomodullin may be purified. In this case, by monitoring an increase in the cAMP activity of platelets, a fraction containing AM of interest can be obtained. Methods for isolation and purification of AM are described in Japanese Laid-Open Publication No. 7-196693.

When AM is produced using recombinant DNA techniques, the DNA sequence encoding a peptide of interest is expressed using various recombinant systems. Construction of expression vectors and preparation of transformants having appropriate DNA sequences are conducted by methods known in the art. Expression may be conducted using prokaryote systems or eukaryote systems.

Prokaryote hosts used include E.coli, bacillus, and other bacteria. For such prokaryote hosts, plasmid vectors

having replication sites and control sequences compatible with the hosts are used. For example, E.coli is typically transformed with a derivative of pBR322 which is a plasmid derived from E.coli. In this case, the control sequence herein includes a promoter for initiation of transcription, an operator if necessary, and a ribosome binding site. Such a control sequence includes generally used promoters such as for example β -lactamase and lactose promoter systems (Reference 12), tryptophan promoters (Reference 13), and P_L promoters derived from λ and N-gene ribosome binding sites (Reference 14).

As an eukaryote host, yeast and mammalian cells may be used, for example. For such an eukaryote host, a plasmid vector having a replication site and a control sequence compatible with the host is used. For example, yeast is transformed with pYEUra3 (Clontech). Other promoter classes useful in a eukaryote host include, for example, promoters for synthesizing a glycolytic enzyme, which include a promoter for 3-phosphoglycerate kinase (Reference 15); a promoter derived from an enolase gene; a promoter derived from a Leu2 gene obtained from YEp13; a promoter derived from metallothionein; an early or late promoter derived from SV40; and other virus promoters such as for example those derived from polyoma virus, adenovirus II, bovine papilloma virus and avian sarcoma virus. A combination of a host cell and an appropriate promoter is known to those skilled in the art and may be appropriately selected if necessary.

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A transformant can be obtained by introducing an expression vector into an appropriate host cell. A desired AM can be obtained by culturing the transformant under

appropriate conditions.

Chemical synthesis of AM may be conducted within a method known in the art. For example, AM may be synthesized by such a method as a solid phase method on a peptide synthesizer. A C-terminal amidated peptide can be synthesized using a peptide synthesizer by condensing amino acids sequentially from the C-terminal amino acid to the N-terminal amino acid using a benzhydryl amine resin and a standard DCC/HOBt, and cutting out an intended peptide from the resultant peptide resin by a standard cleavage method (trifluoromethanesulfonic acid method).

A C-terminal amidated AM may be obtained by one of the following: a carboxyl group at the C-terminus of the peptide obtained by expression in a host is chemically amidated; or a peptide is prepared so as to have Gly added to the C-terminus of an intended amino acid sequence, and is then allowed to react with the above-mentioned C-terminal amidating enzyme for amidation.

Alternatively, the peptide obtained by adding Gly to the C-terminus of AM may be amidated due to an action of a C-terminal amidating enzyme in vivo as described above.

A disulfide bond can be formed, for example, by oxidizing a peptide by air oxidization or with an appropriate oxidant. The substitution of the disulfide bond can be conducted with a $-\text{CH}_2-\text{CH}_2-$ bond by a well-known method (Reference 16). Generally, cleavage in the disulfide bond is avoided by substituting a $-\text{CH}_2-\text{CH}_2-$ bond for the disulfide bond, resulting in stabilization of the protein.

Assay methods for action of selectively inhibiting myometrial contraction, which are known in the art, may be used to confirm that the thus-obtained AM has an action of inhibiting, preferably selectively inhibiting, myometrial contraction. Examples of such assay methods include: a method using the uterus of a female rat previously treated with estrogen; a method using the uterus of a virgin female rat in proestrus or estrus; a method using the uterus of a female rat during pregnancy or during or after parturition; and the like. When the uterus of a female rat previously treated with estrogen is used, action of inhibiting myometrial contraction may be assayed, for example, under the following conditions: the uterus is isolated from a female rat to which estrogen (for example, 17 β -estradiol) has been administered, and is cut into several parts to obtain uterus fragments. Portions of the uterine strips to which blood vessels are attached are removed to obtain uterine strips. While the resultant uterine strips are immersed in a buffer solution such as for example Ringer's solution, a measurement apparatus, such as for example an isometric transducer and an isotonic transducer, is used to continuously examine myometrial contraction. When a rhythm of uterine muscle in spontaneous contraction become constant, or after bradykinin, oxytocin, or prostaglandin F_{2 α} is added to the solution and thereafter a subject peptide is added to the solution, a change in myometrial contraction is examined. Uterine muscle is caused to contract in the presence or absence of the subject peptide to compare levels of contraction, thereby judging the myometrial contraction inhibiting action of the peptide. In this manner, an action of the subject peptide on spontaneous myometrial contraction, or myometrial contraction induced by bradykinin, oxytocin, or

prostaglandin F_{2α} is determined. If a subject peptide inhibits spontaneous myometrial contraction and bradykinin-induced contraction, but not oxytocin- or prostaglandin F_{2α}-induced myometrial contraction, it is judged that the peptide has a selective myometrial contraction inhibiting action.

III. Preparation of a composition for inhibiting myometrial contraction

A composition of the present invention comprises, in addition to an effective amount of AM, any excipient known to those skilled in the art. Examples of the excipients include lactose, cornstarch, magnesium stearate, and alum.

The composition of the present invention is prepared in accordance with methods known in the art.

The composition of the present invention may be in any form. The composition of the present invention may be a solid, such as for example a tablet, a pill, a capsule, and a granule; or a liquid, such as for example an aqueous solution and a suspension. When the composition of the present invention is orally administered as a tablet, an excipient, such as for example lactose, cornstarch, and magnesium stearate, may be typically used. When the composition of the present invention is orally administered as a capsule, an excipient, such as for example lactose and dried cornstarch, may be typically used. In order to orally administer AM as an aqueous suspension, the AM may be used in combination with an emulsion or a suspension. The aqueous suspension may optionally contain a sweetner and an aroma chemical. When the composition of the present invention is intramuscularly, intraperitoneally,

subcutaneously, or intravenously injected, AM is dissolved in a sterilized solution to prepare a buffer solution which is in turn adjusted into an appropriate pH. When the composition of the present invention is intravenously administered, the composition is preferably isotonic.

The composition of the present invention may be used as an drug for preventing premature labor or miscarriage.

IV. Administration of a composition for inhibiting myometrial contraction

The composition of the present invention may be administered in the form of a conventional peptide formulation as described in Remington's Pharmaceutical Sciences, Mack Publishing, Easton, PA. For example, the composition of the present invention may be administered orally, or alternatively parenterally, such as for example intravenous administration, intramuscular injection, intraperitoneal injection, and subcutaneous injection. These peptides may be supplemented into the amniotic fluid. Preferably, these peptides may also be administered by injection.

When the composition of the present invention is administered into a human, typically, the dose per day can be appropriately determined by those skilled in the art by taking into consideration a patient's symptoms, severity, individual differences in sensitivity, weight, age, and the like. The composition of the present invention may be administered once a day or several times a day.

Premature labor or miscarriage would be prevented by administration of the composition of the present

invention.

(Examples)

Hereinafter, action of AM as an drug of the present invention for inhibiting, or preferably selectively inhibiting, spontaneous myometrial contraction or bradykinin-induced contraction will be more specifically described. The present invention is not limited to the following examples. AM used in the examples is a synthesized peptide consisting of an amino acid sequence from Tyr in position 1 to Tyr in position 50 of SEQ ID NO: 6 (available from Peptide Institute, Inc.).

(Example 1: Effect of AM on spontaneous contraction of the uterus of a female rat)

1 µg of 17 β -estradiol in 0.2 ml of 30% ethanol was subcutaneously injected to 10 to 12 week old female rats.

On the following day, the rats were sacrificed by hammering their heads. Thereafter, the rats were decapitated, followed by exsanguination. The uteri were isolated from the rats. Each isolated uterus was cut into four fragments a to d (Figure 1(A)). Portions of each strip to which blood vessels were attached were cut off, thereby obtaining uterine strips (Figure 1(B)).

The effect of AM on the rat uterus was examined by measuring contractions of the uterine strips using an isotonic transducer TD-112S (manufactured by Nippon Kohden Corporation) where the tension was 1 g.

While the uterine strip was immersed in 30 ml of a modified Krebs-Ringer bicarbonate (KRB) solution with

glucose) (hereinafter simply referred to as "modified KRB solution"), the uterine strip was attached to the isotonic transducer. The composition of the modified KRB solution is as follows: 122 mM NaCl, 26 mM NaHCO₃, 5mM KCl, 1 mM MgSO₄·7H₂O, 0.03 mM EDTA-2Na, 2.4 mM CaCl₂, and 11 mM glucose; pH 7.4).

Myometrial contraction was continuously measured. After spontaneous rhythm of the uterine muscle was confirmed to be constant, 30 μ l of 10⁻⁴ M AM (experimental sample) or distilled water (control sample) was added to the respective modified KRB solution with glucose, where the concentration of AM was 100 nM. 30 minutes after the addition of AM or distilled water, 300 μ l of 4.5 M KCl was added to make a KCl concentration of 45 mM.

The results are shown in Figures 2(a) to 2(d). Here, Figures 2(a) to 2(d) correspond to the results obtained by using the uterine strips for the portions a to d in Figures 1(A). Figures 2(a) and 2(b) show controls. Figures 2(c) and 2(d) show the results of the addition of 100 nM AM. Arrows on the left side of each Figure indicate times at which distilled water or AM was added. Arrows on the right side of each Figure indicate the times at which 45 mM KCl was added.

As shown in Figures 2(a) and 2(b), spontaneous myometrial contraction was not affected by the addition of distilled water. In the case of the addition of AM, spontaneous myometrial contraction was significantly inhibited (Figures 2(c) and 2(d)). Further, since the addition of 45 mM KCl caused strong contradiction both in the control added sample and the AM added sample), it was

found that the addition of AM does not affect muscle contraction due to activation of voltage-dependent Ca channels caused by depolarization of smooth muscle cells in the uterus.

5

Note that when the isometric transducer was used to conduct the same experiment, the same results as above were obtained (data not shown).

10

(Example 2: Concentration-dependent effect of AM on the uterus of a female rat)

15

In a manner similar to that of Example 1, uterine strips were prepared, and attached to an isotonic transducer in a modified KRB solution. Myometrial contraction was continuously measured. After it was confirmed that the spontaneous rhythm of uterine muscle became constant, 30 μ l of 1×10^{-6} , 2×10^{-6} , 7×10^{-6} , 2×10^{-5} , or 7×10^{-5} M AM (experimental samples), or distilled water (control sample) was added to the modified KRB solution at time 0 min (initial addition). 5 min later, 12 min later, 22 min later, and 32 min later, where AM concentrations were 1, 3, 10, 30, and 100 nM, respectively. Thereafter, after 45 minutes from the initial addition of AM or distilled water, 300 μ l of 4.5 M KCl was added to the solution to be a KCl concentration of 45 mM.

25

The results are shown in Figures 3(a) to 3(c). Here, Figures 3(a) to 3(c) correspond to the results obtained by using the uterine strips for the portions b to d in Figures 1(A), respectively. Figures 3(a) and 3(b) show the results of AM addition having various concentrations from 1 to 100 nM. Figures 3(c) shows the results of the controls. Arrows in each figure indicate times at which AM, distilled

30

water, or KCl was added.

As shown in Figures 3(a) and (b), it was found that spontaneous myometrial contraction was inhibited by the addition of AM in a concentration-dependent manner.

(Example 3: Inhibition of bradykinin-induced contraction by AM)

In a manner similar to that of Example 1, uterine strips were prepared and attached to an isotonic transducer in a modified KRB solution. At the time when 10 nM bradykinin (Peptide Institute, Inc.) was added to the modified KRB solution, myometrial contraction was continuously measured. 20 minutes after the addition of bradykinin, 100 nM AM or distilled water was further added.

The results are shown in Figures 4(a) and 4(b). Here, Figures 4(a) and 4(b) correspond to the results obtained by using the uterine strips for the portions a and c in Figures 1(A), respectively. Figure 4(a) shows the results of addition of 100 nM AM. Figure 4(b) shows the results of addition of distilled water. Arrows in each figure indicate times at which bradykinin, AM, or distilled water were added.

As shown in Figures 4(a) and 4(b), bradykinin-induced contraction was inhibited by the addition of AM.

(Example 4: Effect of AM on contraction induced by oxytocin or prostaglandin F_{2α})

8 to 12 weeks old female rats were used to prepare uterine strips in a manner similar to that of Example 1.

Thereafter, the uterine strip was placed in a tissue chamber filled with 30 ml of a modified KRB solution with aeration of 95% O₂/5% CO₂ at 37°C to measure contraction of the uterine strip in a manner similar to that of Example 1. 5 After 40 minutes of equilibrium, the uterine strip was preincubated for 15 minutes in the presence or absence of 1 μM AM[22-52] or 1 μM CGRP[8-37]. Thereafter, AM was added to the modified KRB solution in the tissue chamber to gradually increase the AM concentration from 1 nM to 100 nM, 10 exposing the uterine strip to AM.

In another experiment, rats without injection of 17β-estradiol were used. The effect of 100 nM AM on uterine contraction induced by 10 nM bradykinin, 1 nM oxytocin, or 15 1 μM PGF_{2α} was tested in the presence or absence of 1 μM AM[22-52] or 1 μM CGRP[8-37].

In all measurements, the uterine strips were eventually contracted due to 45 mM KCl depolarization to confirm responses of the myometrium. The results are shown 20 in Figures 6(a) to 6(f) and Figures 7(a) to 7(e).

The uterine strips isolated from the rats treated with 17β-estradiol, a type of estrogen, contracted spontaneously in a rhythmical manner (Figure 6(a); in this case, distilled water was added instead of an AM solution). 25 AM was added to the chamber so as to gradually increase the concentration (1 to 100 nM). Spontaneous contraction was inhibited in a concentration-dependent manner (IC₅₀=23 nM) (Figures 6(b) and 6(c)). The inhibitory effect of AM could be reversed by removal of AM by washing out and exchanging the modified KRB solution even when uterine muscle was 30 completely relaxed with 100 nM AM (Figure 6(c)). The

5 preliminary addition of 1 μ M AM[22-52] or 1 μ M CGRP[8-37] per se had no effect, but it substantially completely prevents the contraction inhibitory effect of the addition of 1 to 100 nM AM (Figure 6(d) and 6(e)). Figure 6(f) is a graph showing comparison of the results shown in Figures 6(b), 6(d), and 6(e).

10 When uterine strips were prepared from rats which were not treated with estrogen, these were spontaneously contracted with various intervals and amplitudes. As shown in Figures 7(a), 7(b) and 7(c), either 1 nM oxytocin, 1 μ M PGF_{2 α} , or 10 nM bradykinin all significantly stimulated contraction. 100 nM AM substantially had almost no effect on contraction induced by oxytocin (Figure 7(a)) or PGF_{2 α} (Figure 7(b)). In the other hand, bradykinin-induced contraction was completely blocked by 100 nM AM which is a concentration where spontaneous contraction can be completely inhibited (Figure 7(c)). The inhibitory effect of AM on bradykinin-induced contraction was eliminated by the preliminary addition of AM[22-52] or CGRP[8-37] (Figures 7(d) and 7(e)).

(Discussion of Examples)

25 To the inventor's knowledge, the above-described examples are the first to demonstrate that AM inhibits autonomous and spontaneous contraction of the uterus of a rat in concentration-dependent and reversible manners. Further, AM inhibited bradykinin-induced contraction, but had no effect on contraction induced by oxytocin or PGF_{2 α} , or contraction caused by a high K stimulus. This suggests that the AM action does not directly relax smooth muscle of the uterus, and selectively inhibits mechanisms generating spontaneous contraction or bradykinin-induced

contraction.

In the above-described examples, the AM uterine contraction inhibiting action was blocked by both AM[22-52] which is an antagonist for an AM receptor and CGRP[8-37] which is an antagonist for a CGRP receptor. Therefore, the AM action is considered to be expressed through the AM receptor and the CGRP receptor. As to that the AM action is blocked by CGRP[8-37], besides the examples, it has been reported that the vasodilatory action of AM was blocked by CGRP[8-37] in the isolated rat mesentery vasculature (Reference 17), that elevation of heart rate and blood pressure due to AM administration into a rat cerebral ventricle was blocked by AM[22-52] or CGRP[8-37] (Reference 18), and the like. Further, it has been reported that in a binding test using the uterus of a rat, AM could display both ¹²⁵I-AM binding and ¹²⁵I-CGRP binding, i.e., AM could bind to not only a binding site for AM but also a binding site for CGRP (Reference 7). These findings support the results obtained in the examples.

Expression of the AM protein or the AM gene in the uterus is as abundant as the expression level in the suprarenal medulla at which AM was discovered (Reference 7 and 3). It has been reported that in rat and human uteruses, AM is expressed in endometrial tissues rather than smooth muscle tissues of the uterus. Therefore, it is inferred that AM produced in a endometrium acts on uterine smooth muscle as a paracrine factor.

30

Further, the clinical importance of the uterine contraction inhibiting action of AM will be discussed.

In the pregnant uterus, an expression amount of AM is increased by a factor of about 1.8 to about 4.5 compared to the non-pregnant uterus. An amount of ^{125}I -AM binding is increased by a factor of about 10, and an amount of ^{125}I -CGRP binding is increased by a factor of about 4 (Reference 7; Reference 4; and Reference 19). However, it has been reported that the expression of CGRP is reduced to the limit of detection or less (Reference 7). Based on these findings and the results of the examples, i.e., "AM inhibits uterine contraction through the AM receptor and the CGRP receptor", a possibility is suggested that in the pregnant uterus, an increased expression amount of AM inhibits uterine contraction, thereby playing an important role in maintaining pregnancy.

Furthermore, in the examples, AM inhibited bradykinin-induced contraction, but not contraction induced by oxytocin and PGF_{2α}. Generally, it is believed that contractions induced by oxytocin and PGF_{2α} play an important role in parturition. While the physiological or pathophysiological importance of bradykinin-induced contractions has not yet been revealed, since bradykinin is inherently an inflammatory mediator locally produced by an inflammation reaction (Reference 20), the possibility has been suggested that an abnormal increase in bradykinin in the pregnant uterus leads to premature labor and miscarriage (Reference 1). Therefore, a possibility is suggested that AM selectively inhibits abnormal contractions due to bradykinin but not contractions during normal parturition due to oxytocin and PGF_{2α}, to prevent premature labor and miscarriage and maintain pregnancy.

The above-described results will be briefly

described below. In uterine strips isolated from a nonpregnant rat, AM inhibited spontaneous rhythmic contraction in a concentration-dependent manner ($IC_{50}=23$ nM). The inhibitory effect of AM was perfectly blocked by either calcitonin gene-related peptide[8-37] (CGRP[8-37]) which is a putative antagonist for the CGRP receptor or AM[22-52] which is a putative antagonist for the AM receptor. AM also attenuated uterine contraction induced by bradykinin. Bradykinin-induced uterine contraction is blocked by either CGRP[8-37] or AM[22-52]. AM do not have an inhibitory effect or contractile responses induced by oxytocin or prostaglandin $F_{2\alpha}$. These results show that AM selectively inhibits spontaneous myometrial contraction and bradykinin-induced contraction.

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10

INDUSTRIAL APPLICABILITY

15

The present invention provides a composition for inhibiting, preferably selectively inhibiting, spontaneous myometrial contraction and bradykinin-induced contraction, comprising AM. This composition is useful in prevention of premature labor and miscarriage, arresting of parturition during cesarean section, and treatment of dysmenorrhea.

CLAIMS

1. A composition for inhibiting spontaneous myometrial contraction or bradykinin-induced contraction, comprising adrenomedullin.

5 2. A composition according to claim 1, used for selectively inhibiting spontaneous myometrial contraction or bradykinin-induced contraction.

10 3. A composition according to claim 1 or 2, used to prevent premature labor.

15 4. A composition according to claim 1 or 2, used to prevent miscarriage.

20 5. A composition according to claim 1 or 2, used to arrest parturition prior to cesarean section.

25 6. A composition according to claim 1 or 2, used to treat dysmenorrhea.

7. A composition according to claim 1 or 2, wherein the adrenomedullin is:

25 (a) a peptide comprising an amino acid sequence from Ser in position 13 to Tyr in position 52 of SEQ ID NO: 2 in SEQUENCE LISTING; or

30 (b) a peptide comprising an amino acid sequence having one or several amino acid deleted, substituted, or added in the amino acid sequence (a), and having an action of inhibiting myometrial contraction.

8. A composition according to claim 7, wherein the

adrenomedullin is:

(c) a petptide comprising an amino acid sequence from Tyr in position 1 to Tyr in position 52 of SEQ ID NO: 2 in SEQUENCE LISTING; or

5 (d) a peptide comprising an amino acid sequence having one or several amino acid deleted, substituted, or added in the amino acid sequence (c), and having an action of inhibiting myometrial contraction.

10 9. A composition according to claim 8, wherein the adrenomedullin is:

(e) a petptide comprising an amino acid sequence from Ala in position -73 to Tyr in position 52 of SEQ ID NO: 2 in SEQUENCE LISTING; or

15 (f) a peptide comprising an amino acid sequence having one or several amino acid deleted, substituted, or added in the amino acid sequence (e), and having an action of inhibiting myometrial contraction.

20 10. A composition according to claim 9, wherein the adrenomedullin is:

(g) a petptide comprising an amino acid sequence from Met in position -94 to Leu in position 91 of SEQ ID NO: 2 in SEQUENCE LISTING; or

25 (h) a peptide comprising an amino acid sequence having one or several amino acid deleted, substituted, or added in the amino acid sequence (g), and having an action of inhibiting myometrial contraction.

30 11. A composition according to claim 1 or 2, wherein the C-terminus of the adrenomedullin is amidated.

12. A composition according to claim 1 or 2, wherein Gly

is added to the C-terminus of the adrenomedullin.

5 13. A composition according to claim 1 or 2, wherein in the adrenomedullin, Cys in position 16 and Cys in position 21 of SEQ ID NO: 2 in SEQUENCE LISTING are crosslinked.

10 14. A composition according to claim 13, wherein the crosslink is a disulfide bond.

15 15. A composition according to claim 13, wherein the crosslink is a -CH₂-CH₂- bond.

15 16. A method for preventing premature labor or miscarriage using a composition comprising adrenomedullin.

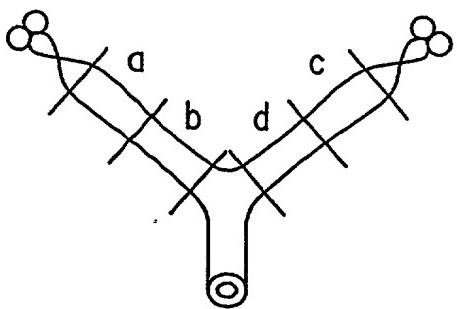
15 17. Use of adrenomedullin in production of a drug for preventing premature labor or miscarriage.

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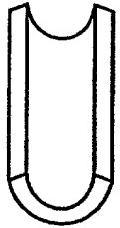
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FIG. 1

(A)



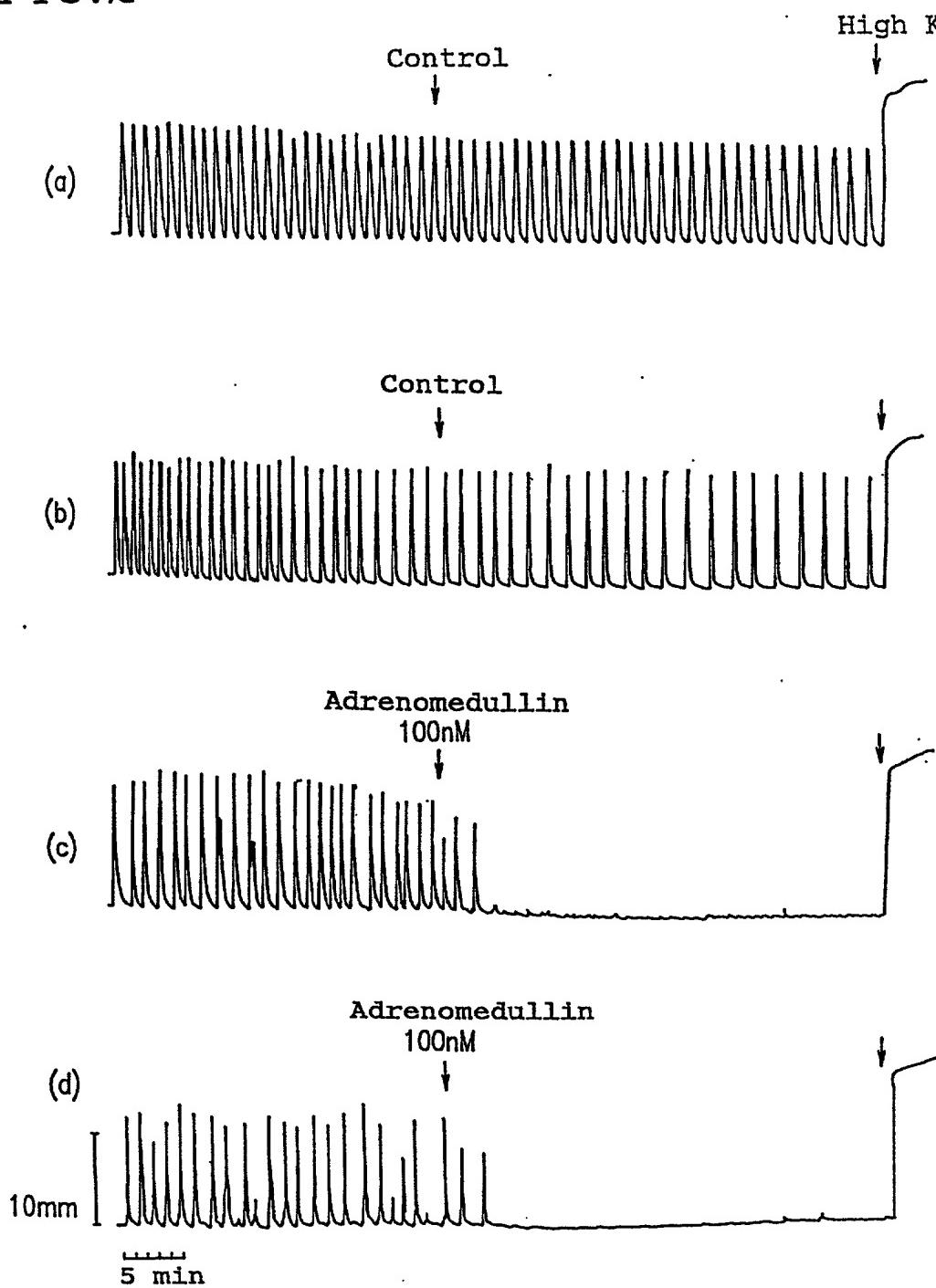
(B)



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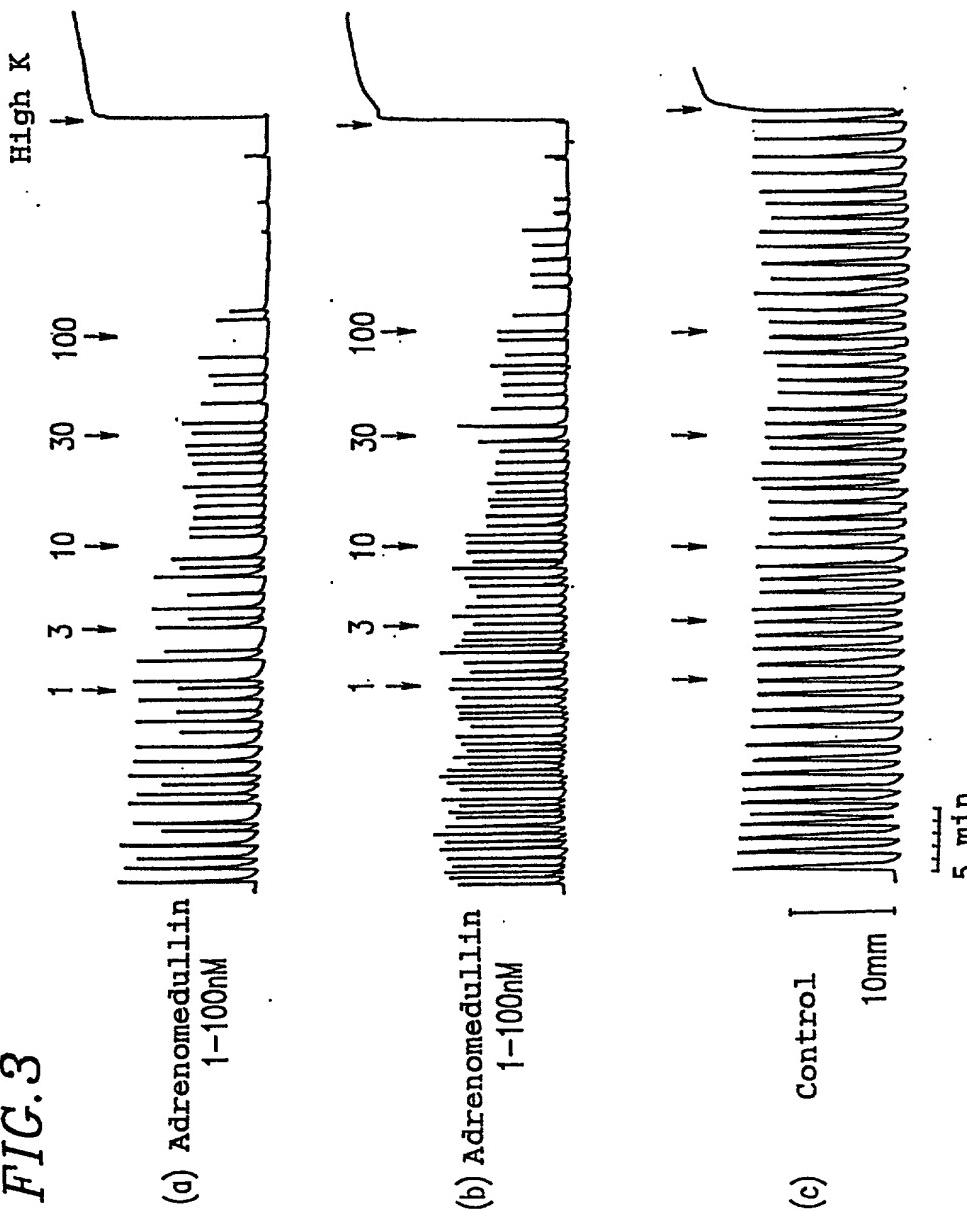
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FIG. 2



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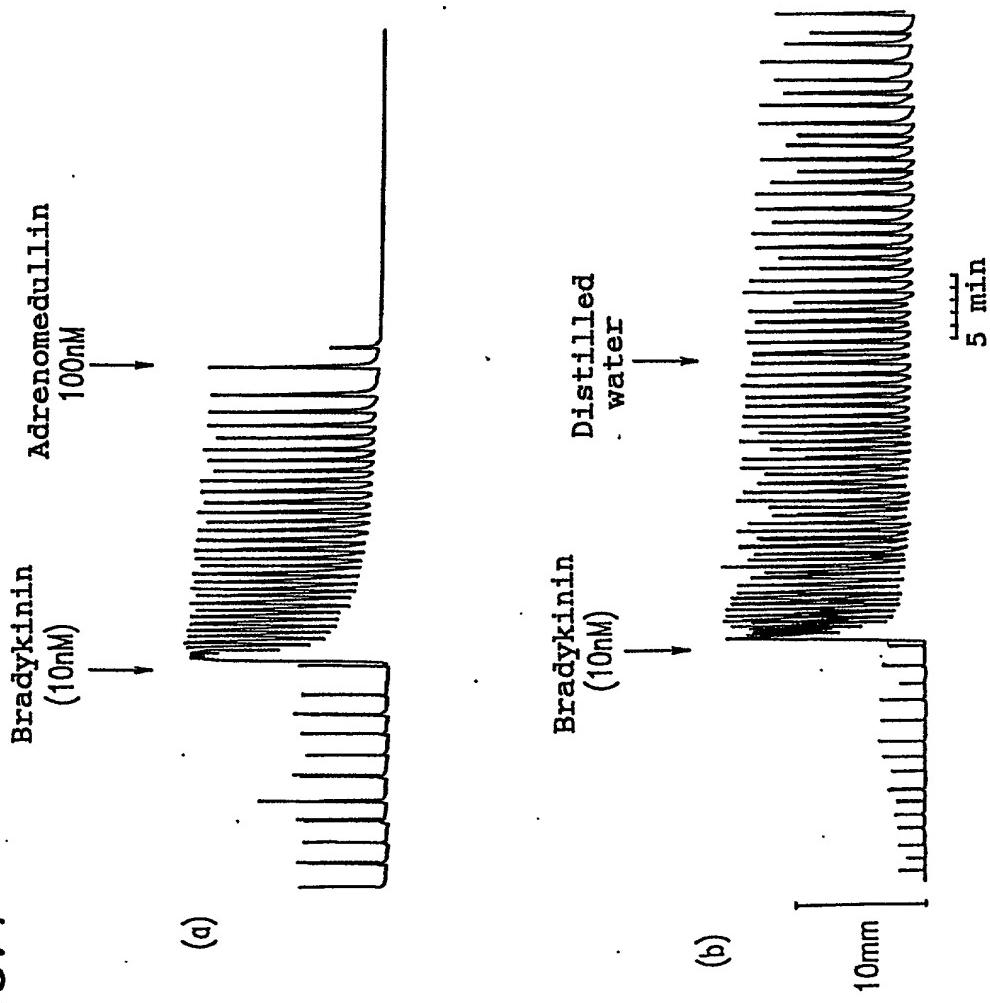
FIG. 3



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FIG. 4



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FIG. 5

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Tyr-Arg-Gln-Ser-Met-Asn-Asn-Phe-Gln-Gly-Leu-Arg-Ser-Phe-Gly-
|RE1| |RE2| |RE3|

10

20
Cys-Arg-Phe-Gly-Thr-Cys-Thr-Val-Gln-Lys-Leu-Ala-His-Gln-Ile-
| | |RE4|

30

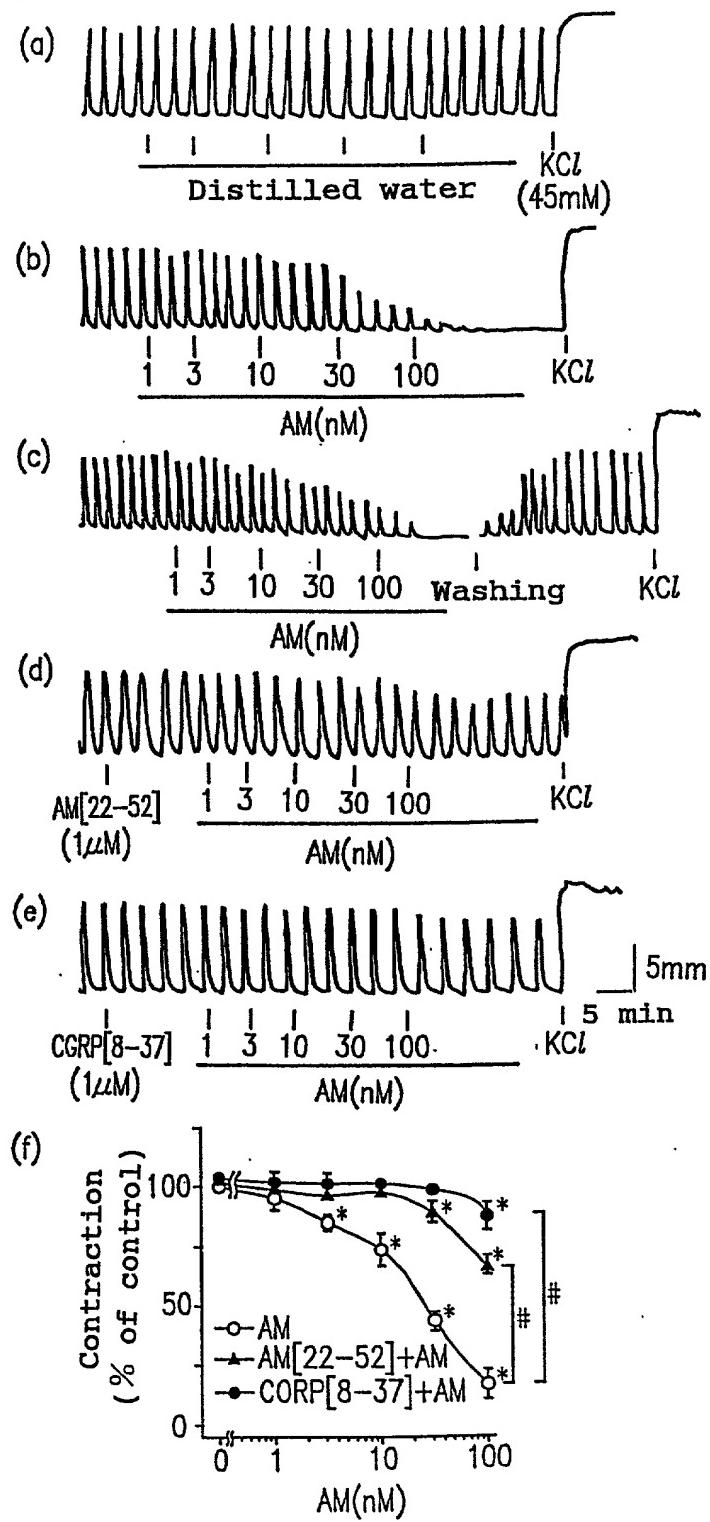
40
Tyr-Gln-Phe-Thr-Asp-Lys-Asp-Lys-Asp-Asn-Val-Ala-Pro-Arg-Ser-
| | | | |RE5|

50 52

Lys-Ile-Ser-Pro-Gln-Gly-Tyr-NH₂
|RE6|

6/7

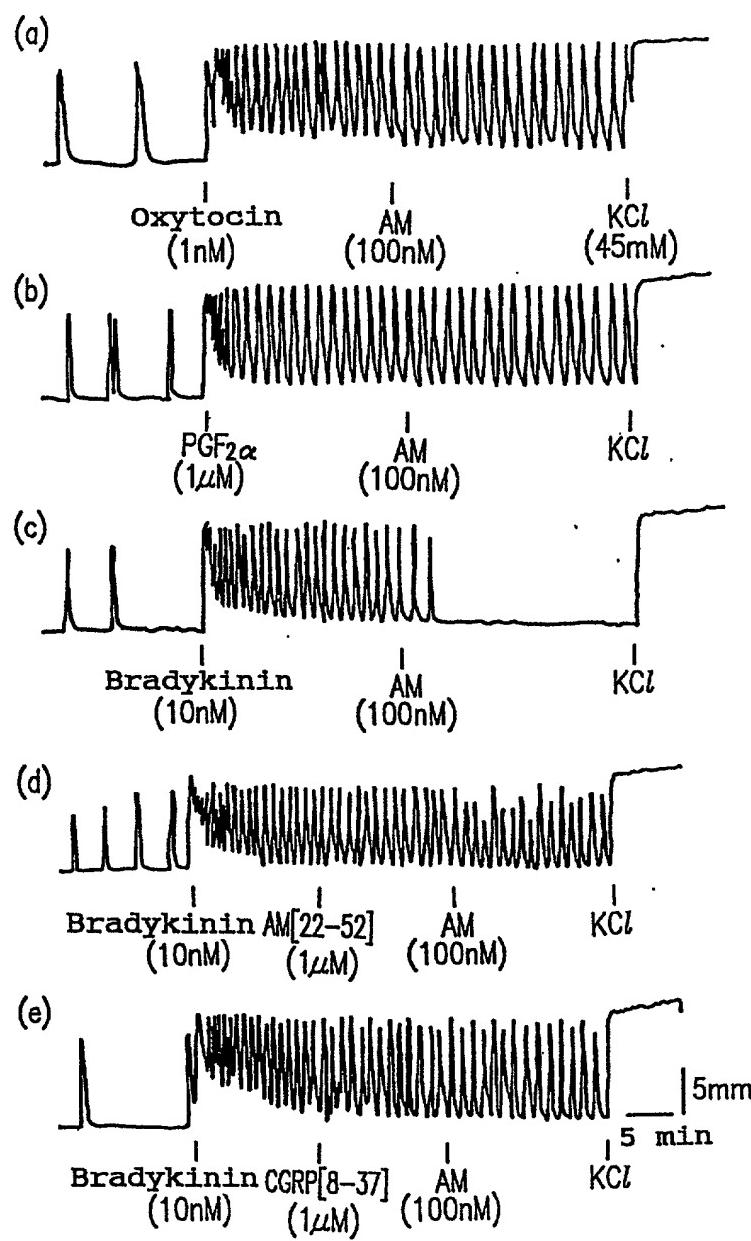
FIG. 6



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FIG. 7



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PATENT APPLICATION
(37 CFR 1.63)**

Declaration Submitted with Initial Filing OR Declaration Submitted after Initial Filing (surcharge (37 CFR 1.16 (e)) required)

Attorney Docket Number	YAM 2 0014
First Named Inventor	Toshihiko YANAGITA
COMPLETE IF KNOWN	
Application Number	/
Filing Date	
Group Art Unit	
Examiner Name	

As a below named Inventor, I hereby declare that:

My residence, post office address, and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

DRUG FOR INHIBITING MYOMETRIAL CONTRACTION

the specification of which

(Title of the Invention)

Is attached hereto

OR

was filed on (MM/DD/YYYY) 06/23/2000 as United States Application Number or PCT International Application Number

(Article 34)
(if applicable)

Application Number PCT/JP00/04167 and was amended on (MM/DD/YY)

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment specifically referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in 37 CFR 1.56.

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Prior Foreign Application Number(s)	Country	Foreign Filing Date (MM/DD/YYYY)	Priority Not Claimed	Certified Copy Attached? YES	Certified Copy Attached? NO
11-177548	JAPAN	06/23/1999	<input type="checkbox"/>	<input type="checkbox"/>	<input checked="" type="checkbox"/>
2000-079171	JAPAN	03/21/2000	<input type="checkbox"/>	<input type="checkbox"/>	<input checked="" type="checkbox"/>

Additional foreign application numbers are listed on a supplemental priority data sheet PTO/SB/02B attached hereto:

I hereby claim the benefit under 35 U.S.C. 119(e) of any United States provisional application(s) listed below.

Application Number(s)	Filing Date (MM/DD/YYYY)

Additional provisional application numbers are listed on a supplemental priority data sheet PTO/SB/02B attached hereto.

[Page 1 of 2]

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DECLARATION — Utility or Design Patent Application

I hereby claim the benefit under 35 U.S.C. 120 of any United States application(s), or 365(c) of any PCT International application designating the United States of America, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C. 112, I acknowledge the duty to disclose information which is material to patentability as defined in 37 CFR 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application.

U.S. Parent Application or PCT Parent Number	Parent Filing Date (MM/DD/YYYY)	Parent Patent Number (If applicable)

Additional U.S. or PCT International application numbers are listed on a supplemental priority data sheet PTO/SB/02B attached hereto.
As a named inventor, I hereby appoint the following registered practitioner(s) to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith: Customer Number →

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Registered practitioner(s) name/registration number listed below

Name	Registration Number	Name	Registration Number
Richard M. Klein	33,000		

Additional registered practitioner(s) named on supplemental Registered Practitioner Information sheet PTO/SB/02C attached hereto.
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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under 18 U.S.C. 1001 and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Name of Sole or First Inventor:		<input type="checkbox"/> A petition has been filed for this unsigned inventor			
Given Name (first and middle if any) Toshihiko		Family Name or Surname YANAGITA			
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<input type="checkbox"/> Additional inventors are being named on the _____ supplemental Additional Inventor(s) sheet(s) PTO/SB/02A attached hereto					